

Temporal Effect of Carbofuran, a Carbamate Insecticide in the Interruption of Estrous Cycle and Follicular Toxicity in Female Swiss Albino Mice

P. N. Baligar, B. B. Kaliwal

Reproductive Toxicology Laboratory, Postgraduate Department of Studies in Zoology, Karnatak University, Dharwad-580 003, India

Received: 26 December 2001/Accepted: 6 May 2003

Carbofuran (2,3-dihydro-2,2-dimethyl-7-benzofuranol N-methyl carbamate), an anticholinesterase carbamate, is commonly used as an insecticide, nematicide and acaricide in agricultural practice around the world (Gupta 1994). The widespread carbofuran use, contaminations has been found in the soil, air, food water, and wild life. Carbofuran is highly toxic to humans and wildlife through the oral and inhalations routes of exposure (Baron 1991). It has been shown that members of this class, such as disulfiram and its metabolite dithiocarbamate, can interfere with catecholamine neurotransmitters metabolism by inhibiting the activity of dopamine- β -hydroxylase (D β H). This is an enzyme that converts dopamine to norepineprine and norepineprine then stimulates the release of GnRH (Maj et al. 1969; Przewlocka et al. 1975; Goldman et al. 1994). Therefore, the present investigation was undertaken to evaluate the temporal effect of carbofuran on estrous cyclicity and follicles in mice.

MATERIALS AND METHODS

Carbofuran (pure 98 %) was kindly provided by Rallis India Ltd., Mumbai and dissolved in olive oil as a vehicle for oral administration. Laboratory bred virgin female Swiss albino mice aged 12–17 weeks, weighing between 24–28 g, showing regular 4–5 days estrous cycle were selected randomly from the breeding stock. The animals were housed in separate cages bedded with paddy husk and had free access to synthetic pellet diet “Gold Mohar” (Hindustan Lever Ltd., Mumbai) and water *ad libitum* throughout the study. The lighting schedule was 12:12 h light and dark cycle at a room temperature $26^{\circ} \pm 1^{\circ}$ C. Animals were divided into 5 groups having 10 animals in each group. In the previous study, 1/5th of the LD₅₀ dose was fixed to 0.7, 1 and 1.3 mg. Treatment with 1.3 mg / kg/d carbofuran seems to be effective dose (Baligar and Kaliwal, 2002). Hence, the dose of 1.3 mg / kg/d was chosen to study the temporal effect of carbofuran on sexual periodicity and follicular dynamics. The Carbofuran was administered by gavage for 5, 10, 20 and 30 days respectively. The control group received an equal volume of olive oil. Daily vaginal smear and body weight were recorded throughout the experiment.

The phases of estrous cycle was determined by observing vaginal smear in the morning (08:00 h to 10:00 h) as described by Cooper et al (1993). Animals were killed by cervical dislocation on day 31st, 24 h after the final exposure. Ovaries of 5 animals in each group were taken for follicular growth studies. The weight of ovaries of the animal nearest to the mean weight of the ovaries of respective group was selected. The

ovaries were fixed in Bouin's fluid, embedded in paraffin, sectioned at 5µm thickness and stained with hematoxylin and eosin. All serial sections of the ovary were counted for various stages of development of follicles as described by Hiremath and kaliwal (2002). Follicles were classified according to Chen et al. (1981) into small, medium and large follicles. Healthy or atretic follicles were classified as described by Swartz and Mall (1989). By using these criteria, mean diameters of follicles have been measured at approximately < 20µm for small, 20-70µm for medium and >70µm for large follicles in mice. Follicles displaying the nucleus of the oocyte were measured by using a calibrated ocular micrometer to avoid repeated counting. The maximum diameter and diameter at the right angle to it were used to obtain a mean diameter for each follicle. A follicle was considered to be undergoing atresia or to regressing whenever two or more pyknotic granulosa cells would be found in a single section or whether the oocyte showed signs of degeneration, such as fragmentation, loss of nuclear membrane, or thinning of cumulus oophorus as proposed by Osman (1985).

The change in the body weight was calculated on the basis of the weight taken 1st day at the time of oral administration was considered as the initial body weight and the weight taken on 31st d before cervical dislocation was considered as the final weight. Ovary, uterus, kidney, adrenal, liver, spleen, thymus and thyroid were dissected out, freed from adherent tissue and weighed to the nearest milligram. To ensure normalization of data for statistical analysis, organs weight were expressed per 100 g body weight. In the preset study treatment with carbofuran in different durational groups, did not alter the weights of uterus, kidney, adrenal, liver, spleen, thymus and thyroid (data not shown). Statistical significance between the control and experimental data were subjected to analysis of variance (ANOVA) together with Dunnett's test ($P < 0.05$).

RESULTS AND DISCUSSION

The results obtained in the present study indicate that the control mice exhibited regular estrous cycle and normal duration of each phases of estrous cycle. Treatment with carbofuran for 30 d caused a significant decrease in the number of estrous cycle and duration of proestrus, estrus and metestrus with a concomitant significant increase in the diestrus phase. However, treatment with carbofuran for 5, 10 and 20 d caused no significant change in the number of estrous cycle and duration of each phases of estrous cycle (Table 1). The members of this class, such as disulfiram and its metabolite dithiocarbamate, can interfere with catecholamine neurotransmitters metabolism by inhibiting the activity of dopamine-β-hydroxylase (DβH). This is an enzyme that converts dopamine to norepineprine and norepineprine than stimulates the release of GnRH. The GnRH release is ultimately affected through the inhibition of DβH (Maj et al. 1969; Przewlocka et al. 1975; Goldman et al. 1994), this could be expected to effect estrous cyclicity.

Plowchalk et al. (1993) have reported that the quantitative assessment of follicle number is an indicator of normal function as well as toxic responses in the ovary. Follicles are the principle functional units of the mammalian ovary. The most important controllers of follicular development are follicle stimulating hormone (FSH) and luteinizing hormone (LH) produced from the pituitary and the ovarian steroid estradiol produced by granulosa cells. Although all follicles are apparently exposed to the same fluctuations in these hormones, not all are equally responsive. Some ovulate and other become atretic, indicating the presence of intragonadal regulatory factors. The

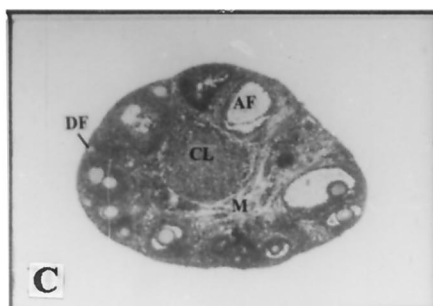
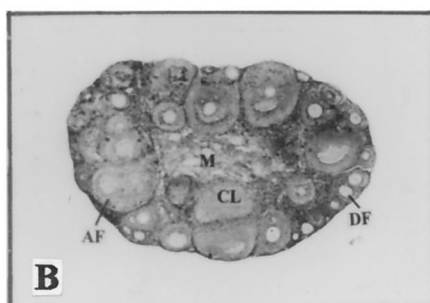
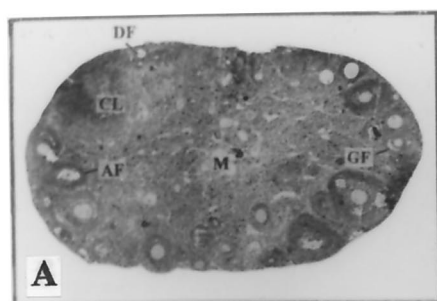


Figure 1. A through C: Temporal effect of carbofuran on ovary.

- A - Section of the ovary of control mice showing the presence of GF, plenty of DF, few AF and many CL.
- B - Section of the mouse ovary treated with carbofuran for days 20, showing the presence of few DF, CL and many AF.
- C - Section of the mouse ovary treated with carbofuran for days 30 showing the presence of few DF, few small CL and many AF.

Abbreviations: GF = Graafian Follicle; DF = Developing Follicle; AF = Atretic Follicle; CL = Corpora Lutea; M = Medulla; (H & E; X 30).

Table 1. Temporal effect of carbofuran on estrous cycle, body and organs weight in albino mice

Groups ^a	Duration of treatment (days)	Number of Cycles	Duration in days (M ± S.E.)				Relative change in body and organ weight (M ± S.E.)	
			Proestrus	Estrus	Metestrus	Diestrus	Body weight (g)	Ovary weight (mg/100g B. Wt)
I	Control	5.9 ± 0.2	4.8 ± 0.3	7.6 ± 0.2	5.5 ± 0.2	11.9 ± 0.4	2.3±0.21	28.81±1.46
II	5	6.1 ± 0.2	4.9 ± 0.4	7.5 ± 0.2	5.3 ± 0.2	11.9 ± 0.5	2.4±0.27	29.98±1.40
III	10	5.7 ± 0.2	4.9± 0.3	7.1 ± 0.2	5.1 ± 0.2	12.3 ± 0.6	2.1±0.28	28.19±1.86
IV	20	5.6 ± 0.3	3.8 ± 0.4	6.9 ± 0.3	4.9 ± 0.3	14.2 ± 0.9	1.7±0.26	27.57±1.11
V	30	4.3 ± 0.3*	1.9 ± 0.3*	5.4 ± 0.3*	3.1 ± 0.2*	19.1 ± 0.5*	1.0±0.26*	18.84±1.88*

A = 10 animals in each group

* = Significant P < 0.05 compared to control

Diestrus Index = $\frac{\text{Number of days with clear diestrus smear}}{\text{Total duration of treatment (days)}} \times 100$

Table 2. Temporal effect of carbofuran on healthy and atretic follicles in albino mice

Groups ^a	Duration of treatment (days)	Number of follicles in the ovary by size classification (μm diameter); mean \pm S.E.						
		Healthy Follicles			Atretic Follicles			
		Small < 20 μm	Medium 20-70 μm	Large > 70 μm	Total	Medium 20-70 μm	Large > 70 μm	Total
I	Control	206.6 \pm 2.7	65.2 \pm 2.4	7.2 \pm 0.4	279.0 \pm 3.0	11.0 \pm 0.9	2.2 \pm 0.4	13.2 \pm 1.1
II	5	205.0 \pm 3.2	65.4 \pm 1.2	7.4 \pm 0.4	277.8 \pm 3.9	11.2 \pm 0.7	2.0 \pm 0.3	13.2 \pm 1.1
III	10	207.6 \pm 4.9	62.4 \pm 1.2	7.0 \pm 0.3	277.0 \pm 5.6	12.0 \pm 0.6	2.4 \pm 0.2	14.4 \pm 0.5
IV	20	201.8 \pm 1.9	60.2 \pm 2.0	6.4 \pm 0.2	269.0 \pm 2.4	12.4 \pm 0.81	2.6 \pm 0.4	15.0 \pm 0.8
V	30	196.2 \pm 2.0*	52.2 \pm 1.9*	4.2 \pm 0.5*	252.6 \pm 2.4*	21.4 \pm 1.1*	3.8 \pm 0.4*	25.2 \pm 0.8*

^a = 5 animals in each group

* = $P < 0.05$ compared to control

histologic observations of the control mice showed normal number of developing follicles, Graafian follicles, corpora lutea and atretic follicles (Fig. A). Histologic observations of the mouse ovary treated with carbofuran for days 5, 10 and 20 showed the presence of developing follicles corpora lutea and many atretic follicles (Fig. B). Treatment with carbofuran for 30 d caused a significant decrease in the number of healthy follicles with concomitant significant increase in the atretic follicles when compared with that of control mice (Table 2). Histologic observation of the mouse ovary treated with carbofuran for 30 d showed the presence of few developing follicles, few small corpora lutea and many atretic follicles (Fig. C). Treatment with carbofuran for days 5, 10 and 20 caused no significant change in the number of different stages of healthy and atretic follicles. This indicates that the effect of carbofuran on estrous cycle and follicular growth is duration dependent. Similar findings have been reported that the administration of number of organophosphate and chlorinated pesticides to adult rats increased the number of atretic follicles with concomitant decrease in the number of some of the follicular stages and total number of healthy follicles (Asmathbanu and Kaliwal 1997; Dhondup and Kaliwal 1997; Math et al. 1998). It has been shown that the ovarian androgen and inhibin secretion by follicles may be an important part in the regulation of FSH secretion and follicular growth (Evans et al. 1997).

In the present study, there is also a possibility that the decreased healthy follicles with concomitant increase in atretic follicles in mice may be due to affecting gonadotropin secretion via central nervous system mechanism, as it was observed in the dithiocarbamates (Goldman et al. 1994). In the present study there is also possibility that the disruption in the estrous cycle, decrease in the healthy follicles with concomitant increase in the atretic follicles may result from damage by toxicant at the level of hypothalamo-pituitary-gonadal axis. However, further investigation in this regard is essential to know the mechanism of action of carbofuran on follicular development and estrous cycle.

Treatment with carbofuran showed a durational related toxicity in terms of body weight (Table 1). There is a significant decrease in the body weight in long-term carbofuran treatment as there may be suppression towards food and water intake. Although food and water intake has not been measured in this study, this may be one of the reasons for low weight gain, which may alter the estrous cycle and folliculogenesis. This finding is important because nutritional deficiency have been shown to alter reproductive function (Simonich and Hites 1995; Hiremath and Kaliwal 2002). The ovary weight was decreased significantly with prolonged carbofuran treatment (Table 1). Similar observations were made in rats treated with monocrotophos and have reported that decrease in weight and size of the ovaries is due to the extensive fibrosis and atretic follicles (Adilaxamma 1994). Therefore, the reasons may be due to the hormonal imbalance in any of the stages in hypothalamo-hypothysial ovarian axis or by insensitising the follicular receptors to the available gonadotropins thereby led to the retardation of further development of surviving follicles into next successive follicular stages and also arrest of estrogen, production which may effect the estrous cycle or directly on the ovary.

Acknowledgement.

We thank U.G.C. New Delhi, for financial support [Grant No. F.3-13/99 (SAP-II)]. Our thanks are due to Prof. S.A. Nevagi, Chairman, Post-Graduate Department of Studies in Zoology, Karnatak University, Dharwad for providing necessary facilities.

REFERENCES

- Adilaxamma K, Janardhan Reddy A, Reddy KS (1994) Monocrotophos: Reproductive toxicity in rats. *Ind J Pharmacol* 26:126-29
- Asmathabanu I, Kaliwal BB (1997) Temporal effect of methyl parathion on ovarian compensatory hypertrophy, follicular dynamics and estrous cycle in hemicastrated albino rats. *Basic and Clinical Physiol and Pharmacol* 8:237-54
- Baron RL (1991) Carbamate insecticides, In: Hayes WJ, Laws ER, (ed.) *Handbook of Pesticide Toxicology*, Academic Press, New York, p. 3-8.
- Chen YT, Mattison DR, Feigenbaum L, Fukui H, Schulman JD (1981) Reduction in oocyte number following prenatal exposure to a diet high in galactose. *Science* 214:1145-7
- Cooper RL, Goldman JM, Vandenbergh JC (1993) Monitoring of the estrous cycle in the laboratory rodent by vaginal lavage. In: Heindel JJ, Chapin RE (ed.). *Methods in Toxicology*, Vol 3 B. San Diego, Academic press, p.45-56
- Dhondup P, Kaliwal BB (1997) Inhibition of ovarian compensatory hypertrophy by the administration of methyl parathion in hemicastrated albino rats. *Reprod Toxicol* 11(1): 77-84
- Dorrington JH, Chuma AV, Bendell JJ (1988) Transforming growth factor and follicle stimulating hormone promote rat granulosa cell proliferation. *Endocrinol* 123:353-59
- Evans ACO, Kumar CM, Wandji SA, Fortune JE (1997) Changes in androgen secretion and leuteinizing hormone pulse amplitude are associated with recruitment and growth of ovarian follicles during luteal phase of the bovine estrous cycle. *Biol Reprod* 57:394-401
- Goldman JM, Stoker TE, Cooper RL, McElory WK, Hein JE (1994) Blocked of ovulation in the rat by fungicide sodium N-methyl dithiocarbamate relationship between effects on the leuteinizing hormone surge and alterations in hypothalamic catecholamines. *Neurotoxicology and Teratology* 16:257-68
- Gupta RC (1994) Carbofuran toxicity. *J Toxicol Environ Health* 43:383-418
- Maj J, Vetulani J (1969) Effect of some N,N-disubstituted dithiocarbamates on catecholamines level in rat brain. *Biochem Pharmacol* 18: 2045-7
- Math JR, Jadaramkunti UC, Kaliwal BB (1998) Effect of edifenphos on follicular dynamics in albino rats. *Indian J Expt Biol* 36:39-42
- Osman P (1985) Rate and course of atresia during follicular development in the adult cycle rat. *J Reprod Fert* 73:261-70
- Plowchalk DR, Smith BJ, Mattison DR (1993) Assessment of toxicity to the ovary using follicle quantitation and morphometrics. In: Heindel JJ, Chapin RE, (ed). *Methods in Toxicology: Female reproductive toxicology*, Vol.3 B. San Diego Academic Press, 57-58
- Przewlocka B, Sarnek J, Szmigielski A, Neiwiakomska A (1975) The effect of some dithiocarbamic acids on dopamine- β -hydroxylase and catecholamines level in rat's brain. *Pol J Pharmacol Pharm* 27:555-9
- Simonich SL, Hites RA (1995) global distribution of Persistent Organochlorine compounds. *Science* 269:1851-1854.
- Soratur SM, Kaliwal BB (1999) Effect of methyl parathion formulation on estrous cycle and reproductive performance in albino rats. *Ind J Expt Biol* 37: 176-8.
- Swartz WJ, Mall GM (1989) Cloredacone induced follicular toxicity in mouse ovaries. *Reprod Toxicol* 3: 203-206.